Stanford TTD: Next-Gen Genomic Imaging

Beckman Center B437, Lokey G3120B



Peter Chou*



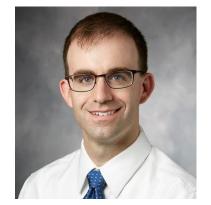
Monica Nagendran*



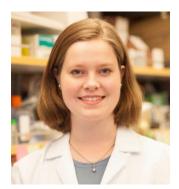
Scott Berger*



Peter Rosston



Adam Andruska



Courtney Stockman



Josh Guild



Jay Mulye



Tushar Desai



P. Harbury

2019 HuBMAP Meeting

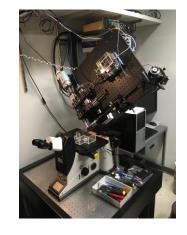
Enable autofluorscence-free 3D imaging

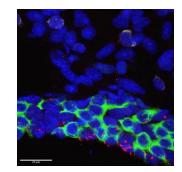
Goals:

- Family of lumiphores that populate color and lifetime channels
- 3D luminescence (time-resolved) microscope
- Rapid electrophoretic *e*Stain and *e*Erase pipeline

Cool results:

- Luminescence light-sheet microscope live
- Deep 3D images with RNA defined cellular shapes





Next year's prospectus

2020 deliverables:

- Accessible luminescence imaging (lumiphores and simple hardware)
- *e*Imaging pipeline and illustrative Ab/RNA mini-maps

Information exchange mechanisms:

- Hosting HuBMAP scientists for hands-on tech transfer
- Workshops?

Electro-SABER collaboration

Activities:

- Six SABER channels set up and tested at Stanford
- Single-site labeling of Fc chains in hamster, rat & human
- RNA mini-map POC complete and probe acquisition underway

Challenges:

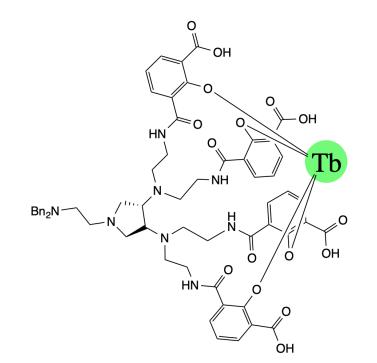
- Time!
- Co-localization of scientists

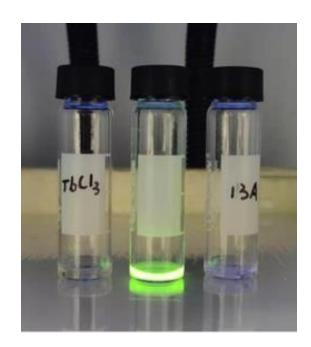
What should HuBMAP do ?

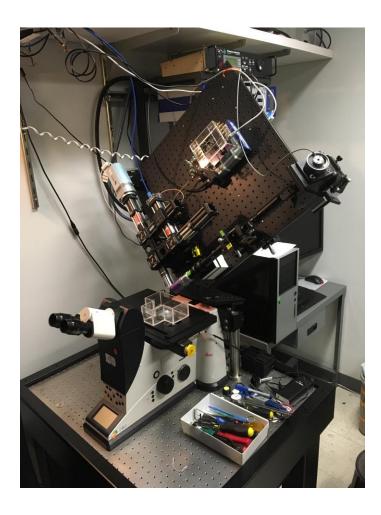
HuBMAP Priorities:

- Cell-type comprehensive spatial maps for human organs with deep RNAseq data
- Easy/intuitive access for bioscience community
- Distribute methods to bioscience community; move beyond cell-type mapping
- Biologist technologist integration

Tissue-optimized luminescence imaging

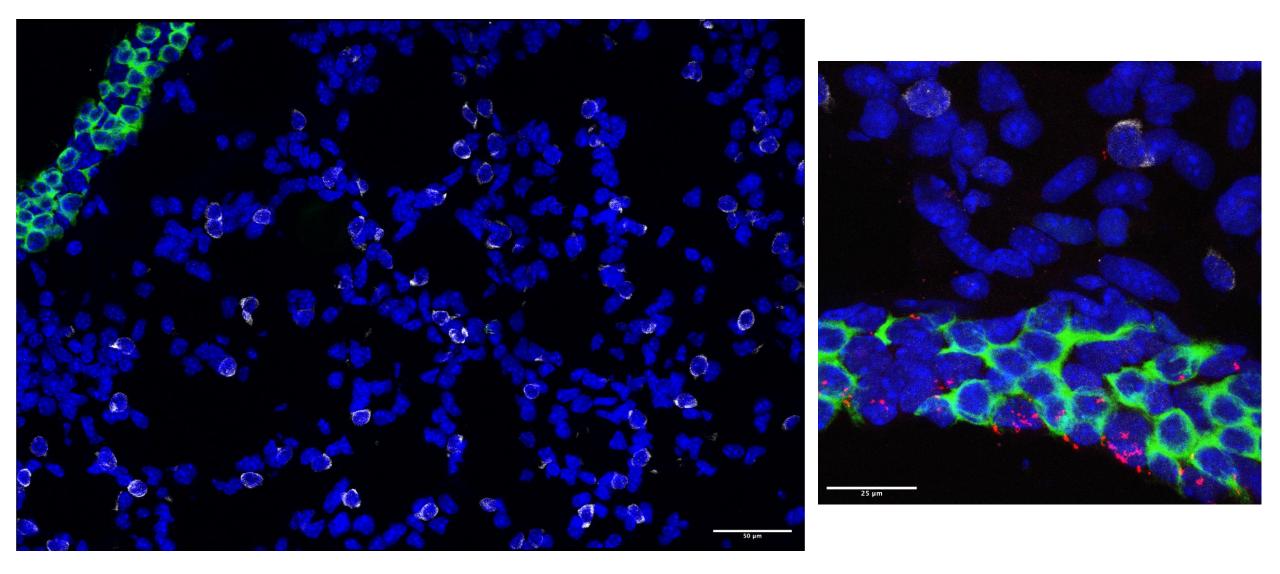




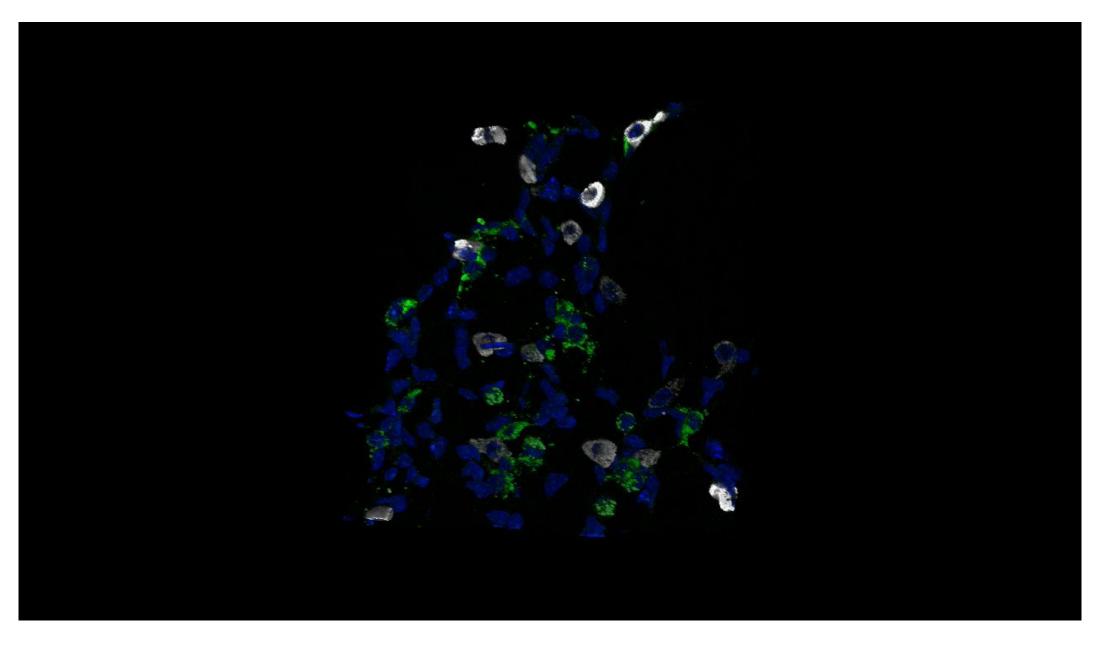


• large axial/lateral FOV (200 um) • simultaneous pan-spectral detection • 14 X 30 X 2.5 mm imaging envelope

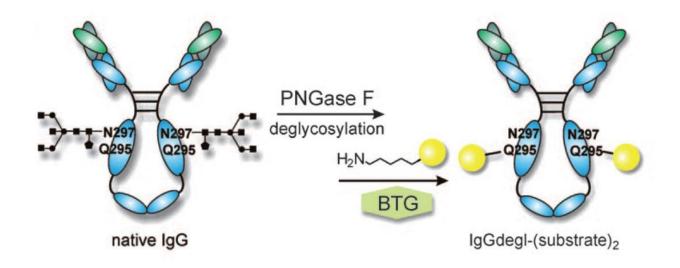
Large tissue blocks (3 X 3 X 3 mm)

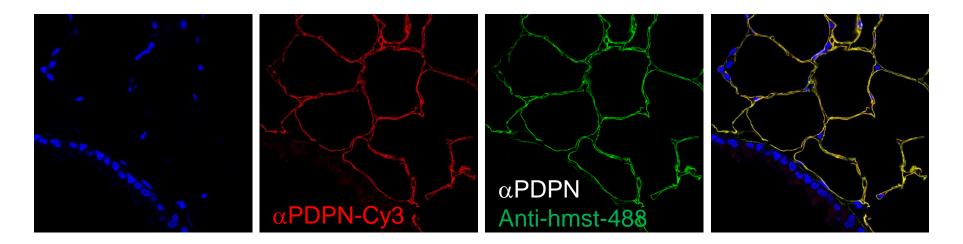


Example 3D data



Enzymatic, site-specific Fc labeling





Jeger et al. Angew. Chem. Int. Ed. 2010 p. 9995